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Cyclopolyamines: Synthesis of Cyclospemidines and Cyclospermine, Analogues of Spermidine and Spermine

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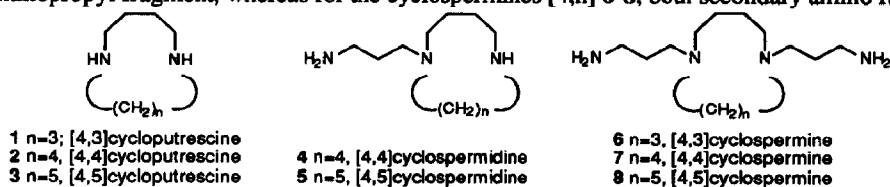
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Abstract: The synthesis of a series of cyclic analogues of the naturally occurring polyamines putrescine, spermidine and spermine was achieved. The cyclic analogues of putrescine named cycloputrescine [4,n] were composed of cycloputrescine in which the two primary amines were connected with a polymethylene chain of variable length ($n = 3, 4, 5$). Cyclospemidines [4,n] and Cyclospermine [4,n] were composed of a cycloputrescine [4,n] moiety mono- or difunctionalized with an aminopropyl chain respectively.

Among various biogenic polyamines, putrescine, spermidine and spermine are ubiquitous in a large variety of biological materials.¹ Although spermine was first isolated from human semen by Leeuwenhoek as early as 1677, as spermine phosphate crystals, only in recent years has much attention been focused on naturally occurring polyamines and their analogues. Since the early 1970s, the literature dealing with these compounds has been grown considerably. Putrescine, spermidine and spermine are found in a wide variety of animals, bacteria, yeasts and plants and are associated with a tremendous variety of biological activity. For example, they have been proposed to stabilise membranes such as *E-coli* spheroplasts², isolated mitochondria³, to facilitate transfection by phage DNA of *E-coli* spheroplasts,⁴ and to modulate membrane functions⁵. They also play important roles in proliferative processes.⁶ In particular, they were shown to be involved in the processes of neoplastic growth^{1,6} as well as, in chemical carcinogenesis.⁷ In this regard, an interesting observation was that in dividing cells, in particular in cancerous cells, the level of polyamines such as putrescine, spermidine and spermine was substantially higher than in other cells.⁸ Based on these observations, the control and the modification of polyamines level was exploited in chemotherapy.⁹ Many polyamines analogues have been prepared and their inhibition effects tested.¹⁰ The control of polyamines level has been also explored by blocking their biosynthesis using D,L- α -difluoromethylornithine (DFMO)¹¹, the most well known specific inhibitor of ornithine decarboxylase.¹²

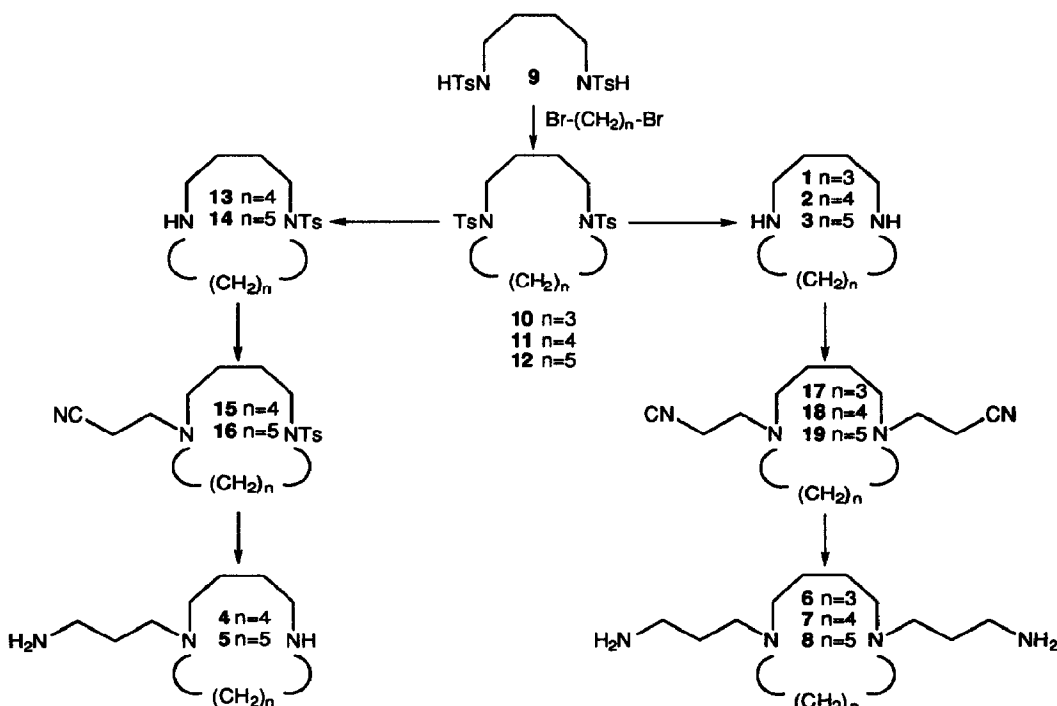
We report here the design and synthesis of a series of spermidine and spermine analogues named cyclospemidine [4,n] and cyclospermine [4,n] respectively. Our design was based on the rigidification of spermidine and spermine structures by interconnection of their two secondary amines with a $-(CH_2)_n-$ ($n = 3, 4, 5$) bridge. Thus, these compounds are all based on a cycloputrescine [4,n] core (1-3). In the case of

cyclospermidines [4,n] **4,5**, their corresponding cycloputrescines [4,n] ($n = 4, 5$) were monofunctionalized with one aminopropyl fragment, whereas for the cyclospermine [4,n] **6-8**, both secondary amino functions of



the cycloputrescines [4,n] ($n = 3, 4, 5$) bear an aminopropyl moiety. Our aim being that these compounds either inhibit the transport of circulant polyamines from outside into the cell, or, if transported, being structural analogues, they may alter cell functions and they may inhibit or at least decrease tumour cells proliferation.

Our strategy for the synthesis of cyclospermidine **4,5** and of cyclospermine **6-8** was based on a stepwise and convergent scheme starting from putrescine. After diprotection of the latter with tosyl group (compound **9**), the medium size cycles **10-12**¹³ were obtained in 8-18 % yield by treatment of the disodium salt of **9** with $\text{Br}-(\text{CH}_2)_n-\text{Br}$; $n = 3-5$ in DMF, following the Richmond and Atkins cyclization procedure.¹⁴ **10-12** were the common intermediates for the synthesis of both cyclospermidines and cyclospermine. Attempts to increase the reaction yield by modification of the cyclization conditions failed. For example the use of K_2CO_3 or Cs_2CO_3 or different addition rate of the dibromo compound did not improve the yield. Since **10-12** are medium size cycles with a ring size of 9 to 11, the results obtained are not surprising. Using the same type of reaction, the effect of the ring size on the yield has been extensively studied by Kellog et al. ¹⁵ Our strategy for the preparation of **4** and **5** required first the synthesis of the monoprotected compounds **13** and **14**.



The synthesis of these two compounds was based on an interesting observation during the deprotection of 10-12 to 1-3 using the standard HBr/AcOH/phenol method.¹⁶ Indeed, the treatment of 11 and 12 by this mixture afforded the monoprotected compounds 13 and 14 in rather high yields.¹⁷ The introduction of the aminopropyl fragment was achieved in two steps by first treatment of 13 or 14 by acrylonitrile¹⁸ in toluene, affording nitriles 15 and 16 in high yields¹⁹, followed by a LiAlH₄ reduction in THF of the cyano group to the primary amine 4 and 5.²⁰ For the synthesis of 6-8, cycloputrescines 1-3 were first prepared by the Li/NH₃ reductive cleavage of the tosyl groups. 1-3 were also previously prepared using many different methods.²¹ The double alkylation of 1-3 by acrylonitrile afforded the dicyano compounds 17-19²² which were transformed into the desired 6-8²³ by treatment with BH₃/THF.

The biological effects of 1-8 as well as their protonation constants will be reported elsewhere.

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References and notes

1. Tabor, C. W., Tabor, H. *Ann. Rev. of Biochem.* **1976**, *45*, 285-306; Smith, T. A. *Endeavour* **1972**, *31*, 22-28; Tabor, H., Tabor, C. W. *Pharmacol. Rev.* **1964**, *16*, 245-300; Seiler, N., Heby, O. *Acta Biochim. Biophys. Hung.* **1988**, *23*, 1-35.
2. Tabor, H., Tabor, C. W. *Adv. Enzymol.* **1972**, *36*, 203-268.
3. Tabor, C. W. *Biochem. Biophys. Res. Commun.* **1960**, *2*, 117-120.
4. Henner, W. D., Kleber, I., Benzinger, R. *J. Virol.* **1973**, *12*, 741-747.
5. Schuber, F. *Biochem. J.*, **1989**, *260*, 1-10.
6. Jänne, J., Poso, H., Raina, A. *Biochem. Biophys. Acta* **1978**, *473*, 241-293; Fillingame, R. H., Jorstad, C. M., Morris, D. R. *Proc. Natl. Acad. Sci. USA* **1975**, *72*, 4042-4045.
7. Scalabrino, G., Poso, H., Hollta, E., Hannonen, P., Kallio, H., Jänne, J. *Int. J. Cancer* **1978**, *21*, 239-245.
8. Flink, I., Pettijohn, D. E. *Nature* **1975**, *253*, 62-63.
9. Morris, D. R., Morton, L. J. in "Polyamines in Biology and Medicine", Dekker, New York, **1981**, pp. 223. Quemener, V., Khan, N. A., Moulinoux, J.-Ph. *The Cancer J.* **1990**, *3*, 45-52.
10. Ganem, B. *Acc. Chem. Res.* **1982**, *15*, 290-298; Bergeron, R. J. *Acc. Chem. Res.* **1986**, *19*, 105-113; Heby, O., Jänne, J. in "Polyamines in Biology and Medicine", D. R. Morris, L. J. Morton eds., Dekker, New York, Basel, **1981**, pp. 243; Sunkara, P. S., Baylin, S. B., Luk, G. D. in "Inhibition of Polyamines Metabolism", P. P. McCann, A. E. Pegg, A. Sjoerdsma, eds., Academic Press, Orlando, **1987**, pp. 121.
11. Bey, P., Schirlin, D. *Tetrahedron Lett.* **1978**, *52*, 5225-5228.
12. Schechter, P. J., Barlow, L. R., Sjoerdsma, A. in "Inhibition of Polyamines Metabolism", P. P. McCann, A. E. Pegg, A. Sjoerdsma, eds., Academic Press, Orlando, **1987**, pp. 345.
13. **General procedure for the synthesis of 10-12.:** In a 1 l flask dry MeOH (400 ml) and Na (2.9 g, 126 mmol) were stirred until dissolution. To the sodium methoxide solution thus obtained, 9 (20 g, 50.4 mmol) was added and the mixture stirred under N₂ at 60 °C for 2 h. before the solvent was removed. To the white solid DMF (400 ml) was added and the mixture stirred under N₂ at 110 °C before the α,ω -dibromo compound (1.1 eq.) was added dropwise (60 min) and the mixture further stirred at 100 °C overnight. The solvent was removed leaving an oil which was dissolved in CH₂Cl₂ (200 ml) and washed successively with H₂O (100 ml), aqueous NaOH (100 ml, 2.5 N) and aqueous HCl (100 ml, 10%). The organic layer was dried (MgSO₄). Filtration over alumina and removal of the solvent left a solid which was recrystallized from hexane/CHCl₃. The crystals obtained were further purified by chromatography (Al₂O₃, CH₂Cl₂/hexane). For all compounds satisfactory elementary analysis on C, H, and N were obtained.
10: 1,3-dibromopropane (7.6 g, 37.6 mmol), yield = 8.5 % (2.6 g). M. P. > 250 °C (decomp.); ¹H-NMR (CDCl₃): 1.87 (br.t, 4H, NCH₂CH₂CH₂CH₂N), 2.21 (q, 2H, NCH₂CH₂CH₂N), 2.44 (s, 6H, CH₃); 3.08-3.16 (br.m, 8H, NCH₂), 7.30-7.35, 7.66-7.69 (m, 8H, arom.); ¹³C-NMR (CDCl₃): 21.6 (CH₃), 21.9, 25.2, 44.7, 50.6 (CH₂), 127.6, 129.8, 134.1, 143.6 (arom.).
11: 1,4-Dibromobutane (10.9 g, 50.5 mmol), yield = 16 % (5.5 g). M. P. > 250 °C (decomp.); ¹H-NMR (CDCl₃): 2.01 (br. q, 8H, NCH₂CH₂); 2.43 (s, 6H, CH₃), 3.19 (br.t, 8H, NCH₂), 7.29-7.33, 7.63-7.66 (m, 8H, arom.); ¹³C-NMR (CDCl₃): 21.6 (CH₃), 24.9, 51.1 (CH₂), 127.6, 129.7, 134.2, 143.4 (arom.).
12: 1,5-Dibromopentane (11.6 g, 50.4 mmol), yield = 14 % (4.9 g). M. P. > 250 °C (decomp.); ¹H-NMR (CDCl₃): 1.58 (br.q, 2H, CH₂CH₂CH₂N); 1.89 (br.m, 8H, NCH₂CH₂), 2.42 (s, 6H, CH₃), 3.08 (br.m, 8H, NCH₂), 7.28-7.32, 7.63-7.67 (m, 8H, arom.); ¹³C-NMR (CDCl₃): 21.6 (CH₃); 24.2, 25.4, 28.5, 51.1, 52.1 (CH₂), 127.4, 129.7, 135.3, 143.3 (arom.).
14. Feigenbaum, A., Lehn, J.-M., *Bull. Soc. Chim. Fr.* **1973**, 198-202; Richman, J. E., Atkins, T. J. *J. Am. Chem. Soc.* **1974**, *96*, 2268-2270.
15. Vriesema, B. K., Buter, J., Kellog, R. M. *J. Org. Chem.* **1984**, *49*, 110-113.
16. Snyder, H.R., Heckert, R.E. *J. Am. Chem. Soc.* **1952**, *74*, 2006-2009.
17. **General procedure for the synthesis of 13 and 14.:** In a 500 ml flask 11 or 12, phenol (2 g), and HBr/AcOH (130 ml, 33 %) were heated to 80 °C for 16 h. The mixture was allowed to cool to r. t. before ether (250 ml) was added and the precipitated hydrobromide salt was filtered and further washed with ether (3x100ml). The coloured solid was passed over a

Dowex anion exchange resin in its basic form (H₂O/EtOH: 50/50) affording the desired pure compound 13 or 14 as white solids.

13: 11 (5.55 g, 12.3 mmol), yield = 68 % (2.5 g). ¹H-NMR (CDCl₃) : 1.19 (br.t, 1H, NH), 1.61 (br.m, 4H, CH₂CH₂NH), 1.77 (br.m, 4H, CH₂CH₂NTs), 2.31 (s, 3H, CH₃), 2.73 (br.m, 4H, CH₂NH); 3.03 (br.t, 4H, CH₂NTs), 7.18, 7.22, 7.52, 7.56 (m, 4H, H arom.); ¹³C-NMR (CDCl₃) : 21.5 (CH₃), 25.5, 26.2, 47.5, 51.0 (CH₂), 127.8, 129.5, 133.5, 143.2 (arom.).

14: 12 (4.9 g, 10.5 mmol), yield = 52% (1.7 g). ¹H-NMR (CDCl₃) : 1.65-1.85 (br.m, 10H, CH₂CH₂CH₂N, CH₂CH₂N), 2.42 (s, 3H, CH₃), 2.82-2.89 (br.m, 4H, CH₂NTs), 3.06-3.12 (br.m, 4H, CH₂NH), 7.28, 7.32, 7.65, 7.69 (m, 4H, H arom.); ¹³C-NMR (CDCl₃) : 21.5 (CH₃), 24.8, 25.5, 26.3, 27.4, 28.0, 48.0, 48.7, 50.0, 50.3 (CH₂), 127.3, 129.6, 135.9, 143.0 (arom.).

18. Rouvier, E., Giacomoni, J.-C., Cambon, A. *Bull. Soc. Chim. Fr.* 1971, 1717-1723.
19. **General procedure for the synthesis of 15 and 16:** In a 250 ml flask, 13 or 14 in dry toluene (80ml) and acrylonitrile (16ml) were refluxed. The reaction was followed by TLC and after 5 days, the liquid was separated and the residue washed with toluene. Washes were combined with the supernatant and the solvent removed affording a solid. The desired compounds 15 and 16 were obtained after chromatography (Al₂O₃, CH₂Cl₂) and were recrystallized from CH₂Cl₂/hexane.
- 15:** 13 (2.2 g, 7.4 mmol), yield 96 % (2.5 g). M. P. = 112 °C. ¹H-NMR (CDCl₃) : 1.56 (br.t, 2H, CH₂CN); 1.86 (br.m, 8H, CH₂CH₂N); 2.42 (s, 3H, CH₃), 2.48 (br.t, 2H, NCH₂CH₂CN), 2.62 (br.t, 4H, CH₂NTs), 3.0 (t, 4H, CH₂N), 7.28, 7.32, 7.60, 7.64 (m, 4H, H arom.); ¹³C-NMR (CDCl₃) : 15.9 (CH₂CN), 21.5 (CH₃), 23.7, 24.6, 49.8, 50.6, 51.1 (CH₂), 119.5 (CN), 127.7, 129.6, 133.4, 143.3 (arom.).
- 16:** 14 (1.4 g, 4.5 mmol), yield 92 % (1.5 g). M. P. = 116 °C. ¹H-NMR (CDCl₃) : 1.55 (br.m, 2H, CH₂CN), 1.69-1.97 (br.m, 10H, CH₂CH₂N), 2.42 (s, 3H, CH₃), 2.46-2.55 (br.m, 2H, NCH₂CH₂CN), 2.74 (t, 4H, CH₂NTs), 3.06 (t, 4H, CH₂N), 7.26, 7.31, 7.64, 7.68 (m, 4H, H arom.); ¹³C-NMR (CDCl₃) : 16.2 (CH₂CN), 21.5 (CH₃), 24.3, 25.1, 25.4, 26.0, 28.2, 51.4, 52.6, 55.0 (CH₂), 119.4 (CN), 127.3, 129.6, 135.7, 143.0 (arom.).
20. **General procedure for the synthesis of 4 and 5:** 15 or 16 was refluxed with 20 eq. of LiAlH₄ in THF for 6 days. 100 eq of NaF was added, the excess of LiAlH₄ was destroyed with a H₂O/THF : 50/50 mixture. The slurry was filtered and the solid washed with hot CHCl₃. The filtrate and washes were combined and evaporated affording an oil which was dissolved in a mixture of MeOH/conc. HCl (50:50, 100 ml) and further washed with CHCl₃ (3x100 ml). The desired hydrochloride salts of cyclosperridines 4 and 5 were obtained as oils after evaporation of the aqueous phase.
- 4:** 15 (0.98 g, 2.88 mmol), yield = 90 % (0.80 g). ¹H-NMR (D₂O) : 1.95-2.15 (br.m, 10H, CH₂CH₂N), 2.72-3.6 (br. m, 12H, CH₂N). ¹³C-NMR (D₂O) : 22.6, 24.1, 36.5, 45.2, 51.8, 52.9, 56.0 (CH₂).
- 5:** 16 (1.1 g, 3.03 mmol), yield = 70 % (0.68 g). ¹H-NMR (D₂O) : 1.83-2.09 (br.m, 12H, CH₂CH₂CH₂N), 3.22-3.50 (br.m, 12H, CH₂N). ¹³C-NMR (D₂O) : 21.9, 23.1, 24.2, 38.9, 47.4, 53.5, 59.7 (CH₂).
21. Stetter, H., Marx, J. *Liebigs Ann. Chem.* 1957, 607, 59-66; Glover, G. I., Smith, R. B., Rapoport, H. *J. Am. Chem. Soc.* 1965, 87, 2003-2011; Yamamoto, H., Maruoka, K. *J. Am. Chem. Soc.* 1981, 103, 4186-4194; Alder, R. W., Eastment, P., Moss, R. E., Sessions, R. B., Stringfellow, M. A. *Tetrahedron Lett.* 1982, 23, 4181-4184; Börjesson, L., Welch, C. J. *Acta Chim. Scand.* 1991, 45, 621-626.
22. **General procedure for the synthesis of 17-19:** In a 250 ml flask, toluene, 1,2 or 3 and acrylonitrile (20 eq.) were refluxed under N₂ for 10 days. The solvent was removed and the solid was taken up in CH₂Cl₂ and washed with H₂O (3x20 ml). The organic layer was dried (MgSO₄) and evaporated to dryness. The desired compounds were obtained by chromatography (Al₂O₃, CH₂Cl₂).
- 17:** 1 (2.5 g, 19.5 mmol), yield = 29 % (1.34 g). ¹H-NMR (CDCl₃) : 1.30 (q, 2H, NCH₂CH₂CH₂N), 1.62 (br.m, 4H, CH₂CH₂N), 2.37 (br.m, 8H, CH₂CH₂CN); 2.61-2.70 (br.m, 8H, CH₂N); ¹³C-NMR (CDCl₃) : 16.8 (CH₂CN), 22.3, 27.5, 46.7, 52.3, 53.5 (CH₂), 119.6 (CN).
- 18:** 2 (1.0 g, 7.8 mmol), yield = 33 % (0.61 g). ¹H-NMR (CDCl₃) : 1.68 (br.m, 8H, CH₂CH₂N), 2.45 (br.m, 4H, CH₂CN), 2.51 (br.m, 4H, CH₂CH₂CN), 2.63 (br.m, 8H, CH₂N); ¹³C-NMR (CDCl₃) : 15.5 (CH₂CN), 24.1, 49.6, 51.6 (CH₂), 119.6 (CN).
- 19:** 3 (1.4 g, 9.0 mmol), yield = 21 % (0.50 g). ¹H-NMR (CDCl₃) : 1.55 (br.m, 10H, CH₂CH₂CH₂N), 2.40 (br.m, 12H, CH₂CN), 2.64 (t, 4H, CH₂CN); ¹³C-NMR (CDCl₃) : 15.7 (CH₂CN), 24.9, 25.9, 26.3, 50.7, 53.6, 54.5 (CH₂), 119.6 (CN).
23. **General procedure for the synthesis of 6-8:** 17, 18 or 19 was refluxed in BH₃/THF (15 eq., 1M) for 2 days. The excess of BH₃ was cautiously destroyed with H₂O/THF : 50/50 mixture before the solvents removed. To the residue a solution of H₂O/MeOH/HCl : 1/6/1 was added and the mixture refluxed for 6 h. The mixture was evaporated to dryness and the residue coevaporated with absolute EtOH before it was taken up in aqueous NaOH (100 ml, 2.5 N) and extracted with CH₂Cl₂. The desired tetraamine was obtained as an oil after removal of the solvent. Compounds 6-8 were stored as their hydrochloride salts.
- 6:** 17 (1.2 g, 5.1 mmol), yield = 70 % (1.4 g). ¹H-NMR (D₂O) : 2.29 (br.m, 10H, CH₂CH₂N); 3.29-3.70 (br.m, 16H, CH₂N). ¹³C-NMR (D₂O) : 22.8, 24.7, 26.1, 39.1, 55.4, 57.0, 58.4 (CH₂).
- 7:** 18 (0.30 g, 1.2 mmol), yield = 90 % (0.45 g). ¹H-NMR (D₂O) 2.26 (br.m, 12H, CH₂CH₂N); 3.35-3.80 (br.m, 16H, CH₂N). ¹³C-NMR (D₂O) : 2.24 (br.m, 14H, CH₂CH₂N); 3.24-3.70 (br.m, 16H, CH₂N); ¹³C-NMR (D₂O) : 26.1, 28.4, 39.7, 53.8, 57.4 (CH₂).
- 8:** 19 (0.50 g, 1.9 mmol), yield = 92 % (0.83 g). ¹H-NMR (D₂O) : 2.24 (br.m, 14H, CH₂CH₂N); 3.24-3.70 (br.m, 16H, CH₂N); ¹³C-NMR (D₂O) : 25.0, 25.9, 26.1, 31.3, 40.7, 52.9, 53.8, 54.7 (CH₂).

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